

The interactive effects of ketamine and magnesium upon depressive-like pathology

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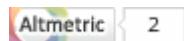
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Abstract: Approximately one-third of patients with major depressive disorders (MDDs) are resistant to current treatment methods, and the majority of cases relapse at some point during therapy. This has resulted in novel treatments being adopted, including subanesthetic doses of ketamine, which affects aberrant neuroplastic circuits, glutamatergic signaling, and the production of brain-derived neurotrophic factor. Ketamine rapidly relieves depressive symptoms in treatment-resistant major depressive disorder patients with effects that last for up to 2 weeks even after a single administration. However, it is also a drug with an abusive potential and can have marked side effects. Hence, this study aimed at enhancing the antidepressant-like effects of ketamine (allowing for lower dosing regimens) by coadministering magnesium hydroaspartate (Mg^{2+} normally affects the same receptors as ketamine) and also assessed whether an Mg^{2+} -deficient diet would modify the impact of ketamine. It was found that a single 15 mg/kg dose of ketamine did indeed induce rapid antidepressant-like effects in the forced swim test but did not affect brain levels of the brain-derived neurotrophic factor. Contrary to our hypothesis, magnesium administration or deficiency did not influence the impact of ketamine on these outcomes. Thus, these data do not support the use of magnesium as an adjunct agent and instead suggest that further research involving other antidepressant and animal models is required to confirm the present findings.

Keywords: ketamine, depression, treatment resistance, ghrelin, BDNF, NMDA

Introduction

Major depressive disorder is a recurrent chronic psychiatric condition that affects over 121 million people worldwide annually.¹ Unfortunately, the currently used antidepressants (even when used in combination) have several caveats, including a slow onset of action, requiring weeks or months of treatment to produce therapeutic effects.^{2,3} Moreover, recent estimates indicate that approximately 30% of depressed patients do not respond to these antidepressant therapies, and even in case of responders, residual symptoms are still often present and there is a very high risk of relapse.⁴⁻⁷

Exciting emerging evidence indicates that the noncompetitive N-methyl-D-aspartate (NMDA) glutamate receptor antagonist, ketamine, which is widely used for its anesthetic and analgesic properties, could be a useful treatment option for depression.⁸⁻¹⁰ When used at low doses, a single injection of ketamine was found to promote fast-acting (within hours) antidepressant and antisuicidal effects.^{8,11-13} Importantly, ketamine was rapidly effective in patients who were previously treatment-resistant.^{3,11,14}

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neurotoxic effects in animal models.²¹ A better potential treatment strategy might involve the coadministration of agents that can synergize with ketamine in the promotion of an antidepressant response. This would allow for even lower doses and a less frequent ketamine-dosing regimen. In this regard, we posit that magnesium (Mg^{2+}) could be a potential cofactor that might augment the effects of ketamine. Indeed, Mg^{2+} is normally present as a channel blocker on the NMDA receptor, upon which ketamine exerts its antagonist actions. Moreover, there are reports indicating that Mg^{2+} is involved in the pathophysiology of mood disorders^{22–24} and that administration of the Mg^{2+} organic salt, magnesium hydroaspartate, created antidepressant-like effects.^{25,26} In fact, elevated intracerebral Mg^{2+} levels were reported with several existing antidepressant agents.^{27,28}

In this study, the potential additive or synergistic antidepressant-like actions of ketamine plus magnesium treatment were explored. In order to ascertain whether any interactive effects of ketamine and magnesium might be related to trophic brain changes, BDNF levels within the hippocampus and PFC were assessed.

Materials and methods

Animals

Male mice (CD1, 6–8 weeks of age) were acclimatized to the vivarium for a period of 1 week before experimental procedures began. Animals were singly housed in standard polypropylene cages (27×21×14 cm), and a 12-hour light/dark cycle (light phase: 8:00 to 20:00 hours) was maintained. Water and Teklad Global mouse chow were provided ad libitum, and the room temperature was maintained at 21°C. All the procedures were approved by the Carleton University Animal Care Committee and adhered to the guidelines of the Canadian Council for the care of animals in research.

Treatment procedures

This experiment examined the effects of chronic administration of magnesium hydroaspartate prior to a single ketamine injection. Thirty-two mice were randomly divided into two groups. ($n=16$ per group): 1) saline and 2) magnesium hydroaspartate. The treatments were administered intraperitoneally once daily for 14 days. Then each group of mice was subdivided into two groups ($n=8$ per group); one receiving ketamine (15 mg/kg) and the other saline on the test day. Injection side was randomized, and sterile procedures were used to reduce the chances of any infection/inflammation.

Behavioral procedure

On the test day, which was 24 hours after the last magnesium injection, mice were injected either with ketamine or with saline. One hour later, animals were subjected to the forced swim test (FST). The FST was performed based on the methods suggested by Liu et al.¹⁵ Mice were placed in a 2,500-mL beaker filled two-thirds with 22°C±1°C water. They were allowed to swim for 6 minutes during which time their activity was videotaped. Immobility was defined as the absence of active, escape-oriented behaviors such as swimming, jumping, rearing, and sniffing and was assessed during the last 240 seconds of the test. The recorded videos were later scored by a person blind to the treatment groups.

In order to assess whether the drug treatments might be affecting the motor activity (which could confound FST results), the open field test was used, as described by Beurel et al.²⁹ A 50 cm³ open, black Plexiglas arena floor was divided into 36 squares (8×8 cm); then each animal was randomly placed in one of the four corners, and their locomotor activity was recorded for a period of 10 minutes. The number of lines crossed by each mouse was counted by a person blind to the study. The open field test was also used to assess anxiety-like behaviors in mice, as previously described by Littelljohn et al.³⁰ The open, black Plexiglas arena was divided into predefined zones (outer, middle, and inner/center). The amount of time spent in each zone was measured by a person blind to the study groups. Decreased time spent in the center zone is an index of anxiety-like behavior. The arena was cleaned with 10% EtOH between trials.

Immediately after testing, all the animals were rapidly decapitated and brains collected. The hippocampus and PFC were carefully microdissected from 0.5 mm coronal tissue slices. All the tissues were placed on dry ice followed by storage at –80°C until assayed.

BDNF analyses

Prior to analysis, samples were removed from the freezer and weighed. Lysis buffer (100 mM PIPES [pH 7], 500 mM NaCl, 0.2% Triton X-100, 0.1% Na₃N, 2% BSA, 2 mM EDTA·Na₂·2H₂O), 200 μM

Phenylmethylsulfonyl fluoride (frozen in isopropanol), 10 μM leupeptin (frozen separately in deionized water), and 0.3 μM pepstatin (frozen separately in dimethyl sulfoxide) were then pipetted into each tube containing hippocampal or PFC samples (2 mL). Samples were then homogenized and sonicated (Virtis Virsonic; Virtis Company, Gardiner, NY; with a microtip at power level 4 and pulses at 1-second intervals for 15 seconds). Then an additional 2 mL of lysis buffer was added to the samples, and they were again sonicated. Samples were split and one half of each sample was spiked (artificially increasing the concentration of BDNF by 250 pg/mL to determine percent recovery). Samples were centrifuged for 30 minutes at 16,000× *g* at 4°C. Supernatants were removed and frozen at –80°C until analysis.

BDNF expression was determined by Western blot analysis. Protein concentration in the supernatants of tissue extracts was determined using a bicinchoninic acid protein assay kit (Pierce Biotechnology, Inc., Rockford, IL, USA). A quantity of 30–50 μg of total proteins was loaded onto a 4%–15% gradient polyacrylamide gel, electrophoretically transferred to polyvinylidene difluoride membrane and probed with the following primary antibody: BDNF (1:1,000; Santa Cruz Biotechnology, Inc., Dallas, TX, USA). β-Actin

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presented in the form of mean \pm standard error of the mean. All data were analyzed using the statistical software SPSS (Version 19.0), and the differences were considered statistically significant when $P < 0.05$.

Results

Forced swim test

As shown in [Figure 1](#), mice treated with ketamine (15 mg/kg) spent significantly less time immobile than their saline-injected counterparts ($F_{(1, 30)} = 5.2, P = 0.031$). However, forced swim immobility was neither affected by the magnesium treatment ($F_{(1, 30)} = 0.686, P = 0.415$) nor was there any indication of an interaction between ketamine and magnesium ($F_{(1, 30)} = 0.873, P = 0.359$).



Figure 1 Immobility in the forced swim test.

Notes: Effect of ketamine and magnesium treatment on immobility time in the forced swim test. Mice treated with ketamine (15 mg/kg) spent significantly less time immobile than the saline group ($*P < 0.05$). No significant effect of ketamine and magnesium treatment on immobility time. A significant main effect for ketamine was obtained by ANOVA. All data are expressed as mean \pm standard error of the mean.

Locomotor activity in the open field test

As shown in [Figure 2](#), the overall locomotion activity was not influenced by ketamine or magnesium treatment ($F_{(1, 32)} = 0.886, P = 0.358$), indicating that the treatments were not having general deleterious consequences on motor behavior.

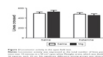


Figure 2 Locomotor activity in the open field test.

Notes: Locomotor activity was expressed as the total number of lines passed by mice over 10 minutes in a 50 cm³ open white Plexiglas arena, which was divided into 36 squares, each 10 cm. No significant difference among groups was observed. All data are expressed as mean \pm standard error of the mean.

Anxiety-like behavior in the open field test

In the open field arena, no significant difference was found between the time spent in the center zone ($F_{(1, 32)} = 0.143, P = 0.708$), middle zone ($F_{(1, 32)} = 0.148, P = 0.734$), or outer zone ($F_{(1, 32)} = 0.066, P = 0.798$; [Figure 3](#)). Importantly, reduced exploration of the central portion of this arena is typically taken as an index of anxiety-like behavior.



Figure 3 Anxiety-like behavior in the open field test.

Notes: Effect of ketamine and magnesium treatment on anxiety-like behavior in the open field test. Time spent in the center zone ($P = 0.708$), middle zone ($P = 0.734$), and outer zone were measured in seconds. No significant difference was found in any of the groups. All data are expressed as mean \pm standard error of the mean.

PFC and hippocampal BDNF expression

Western blot analysis of hippocampal BDNF protein levels revealed no significant ketamine \times magnesium treatment interaction ($F_{(1, 32)} = 0.08, P = 0.90$). The main effects of either magnesium ($F_{(1, 32)} = 1.43, P = 0.241$) or ketamine ($F_{(1, 32)} = 0.186, P = 0.669$; [Figure 4](#)) were also not revealed. Similarly, PFC BDNF protein levels did not vary by ketamine \times magnesium treatment interaction ($F_{(1, 32)} = 1.244, P = 0.356$), and the main effects of magnesium ($F_{(1, 32)} = 0.916, P = 0.347$) or ketamine ($F_{(1, 32)} = 1.46, P = 0.236$) were not significant.

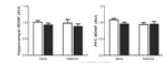


Figure 4 Results of BDNF Western blot.

Notes: Prefrontal cortex BDNF on the right side and hippocampal BDNF on the left side. No significant difference was found among groups either for PFC BDNF or hippocampal BDNF.
Abbreviations: BDNF, brain-derived neurotrophic factor; PFC, prefrontal cortex.

Discussion

Current pharmacotherapies for major depression include selective serotonin and noradrenaline reuptake inhibitors, which have a long delay of action and actually fail to work in many patients.² Therefore, there is an urgent need for the development of faster-acting antidepressants, as well as novel nonmonoaminergic agents to benefit a large number of treatment-resistant patients. In this regard, a lot of recent works have been devoted to examining the antidepressant efficacy of compounds targeting the glutamatergic pathway. This research has been spurred on by the finding that the ionotropic glutamate N-methyl-D-aspartate receptor is involved in the etiology of major depression.^{8,9,19} Since then, numerous clinical and prehuman

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eukaryotic elongation factor 2 kinase, which leads to reduced eukaryotic elongation factor 2 phosphorylation and consequent desuppression of BDNF translation.^{2,20} It also bears mentioning that a recent study³⁶ highlights the role of early and sustained activation of α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid receptors in ketamine's antidepressant responses rather than NMDA receptor inhibition.

While there remains much to be discovered about how ketamine promotes antidepressant outcomes, it is important to keep in mind that a major impetus of this line of inquiry has been the considerable concern over the drug's potential adverse effects (tolerance, abuse, side effects).^{2,19,37} Indeed, the hope is that a better understanding of the molecular and cellular pathways subserving ketamine's antidepressant action will lead to the development of novel glutamatergic drugs that share with ketamine its mode of action/antidepressant potential but not its unwanted effects.³⁸ In the meantime, further studies are needed to find the minimum effective and safe dose of ketamine or to find a favorable ketamine combination therapy that could permit the down-titrating of drug doses.

Poleszak et al³⁹ demonstrated the antidepressant-like effect of magnesium in conjunction with NMDA antagonists (CGP 37849, MK-801, L-701,324) in mice; importantly, these authors suggested that administration of magnesium could be an effective method to reduce the NMDA antagonist dose. Magnesium as an iatrogenic NMDA receptor antagonist has a proven antidepressant-like activity in the FST.^{25,26} Furthermore, it has been used as a supplementary therapy for mood disorders, reducing significantly the use of lithium, benzodiazepines, and neuroleptics in joint therapy.⁴⁰

The present study evaluated the effect of ketamine, delivered either as monotherapy or in combination with magnesium on hippocampal and PFC BDNF expression and behavioral performance. Locomotor activity and anxiety-like behavior in an open field were unaffected by ketamine or magnesium treatment. Similarly, previous studies demonstrated that neither ketamine nor magnesium has any effect on locomotor activity in mice.^{22-24,41-43} However, as expected, we found that ketamine reduced immobility in a FST, and this indicated that the drug had antidepressant-like effects that were unrelated to any anxiety or locomotor parameters.

Magnesium-deficient diets were reported to induce anxiety-related behavior in mice,⁴⁴ and there are studies indicating an anxiolytic-like activity of magnesium in mice.²²⁻²⁴ This, however, contradicts the results of our present study. Furthermore, animal data regarding the impact of ketamine on anxiety behaviors are contradictory: in a study conducted by Silvestre et al,⁴⁵ ketamine was found to produce anxiogenic-like effects in rats, but in another study, the drug was shown to promote anxiolytic responses (in the elevated plus maze).⁴⁶ Similarly, the evidence for an anxiolytic-like effect of ketamine in humans is equivocal and is probably subject to complex dose-and-time effects. For instance, in a study of healthy human volunteers, the participants reported a decrease in anxiety after a low dose of ketamine (0.1 mg/kg intravenously), but an increase in anxiety at a higher dose (0.5 mg/kg intravenously).⁴⁷ Ketamine is often used as a preoperative agent for painful medical procedures for both adults and children, presumably due to its dissociative anesthetic and anxiolytic properties.^{48,49} However, the potential anti-anxiety effects of ketamine may be secondary to sedation and analgesia,⁵⁰ which is particularly evident with doses higher than the subanesthetic ones that are used in this study (usually substantially >30 mg/kg).

Pochwat et al⁵¹ evaluated the effect of three different doses of magnesium (10, 15, and 20 mg/kg) in the chronic mild stress model of depression in rats. They observed an antidepressant-like effect of magnesium at a dose of 15 mg/kg in the sucrose preference test, beginning from the third week of treatment. Moreover, the antidepressant-like behavioral effect of magnesium coincided with reduced brain regional (hippocampus, PFC, and amygdala) expression of proteins related to glutamatergic signaling (including GluN1, GluN2A, and PSD-95).

The underlying mechanism(s) of the rapid antidepressant effect of ketamine is/are not known. However, it has been suggested that the fast-acting behavioral antidepressant-like effect of ketamine is related to the rapid upregulation of BDNF and the consequent potentiation of synaptic plasticity.²⁰ Indeed, acute administration of ketamine was shown to induce increased hippocampal and PFC BDNF levels, which coincided with improved behavioral responses in rodents.^{19,52,53} Yet, we presently failed to detect any effect of ketamine on hippocampal or PFC BDNF levels. This might be due to the fact that we did not use mice with depression-like phenotype in our study, as ketamine was shown to improve decreased BDNF expression in inflammation-induced depressed mice⁵⁴ and learned helplessness rats.^{55,56} Indeed, Zhang et al⁵⁷ found that 8 days after a single injection, ketamine did reverse the stressor-induced reduction of BDNF in the PFC and hippocampus on Day 20 of a daily social defeat stressor paradigm. However, Garcia et al⁵⁸ failed to detect any change in hippocampal BDNF levels after chronic mild unpredictable stress as well. The reason for the discrepancies between the studies is not immediately apparent but could certainly have to do with the obvious stress/testing procedural differences, as well as species/strain differences. In particular, the Garcia et al⁵⁸ study used rats, whereas Zhang et al⁵⁷ used C57BL/6 mice, and we used CD1 mice in our study.

Also, we did not detect any effect of magnesium on hippocampal or PFC BDNF levels. This is in contrast to a study⁵⁹ that found that magnesium is, in fact, capable of increasing BDNF expression and various markers of synaptic plasticity. One of the possible reasons we did not see an effect of magnesium might be the subtherapeutic duration of magnesium administration in our experiment. Indeed, Pochwat et al⁵¹ found changes in the level of proteins related to glutamatergic system in PFC, hippocampus, and amygdala after 3 weeks of magnesium treatment. We chose a 2-week period based on the Poleszak et al²³ study in which they found a reduced immobility time in FST after 2 weeks of magnesium administration. Also, an *in vitro* study conducted by Slutsky et al⁶⁰ suggested that synaptic plasticity is sensitive only to magnesium levels in the

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whether magnesium really has any potential as an adjunctive agent with ketamine or even other more traditional antidepressants, such as fluoxetine or citalopram.

Acknowledgments

This paper includes work based on the master thesis submitted by coauthor Sara Razmjou to Carleton University: <https://curve.carleton.ca/cc1977e4-6b95-4ad3-84c4-42cd6c0cb5a5>.

Disclosure

The authors report no conflicts of interest in this work.

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